

Practical 1: Isolation and Identification of Fungi Species

Introduction

Fungi are multicellular organisms descending from Class Opisthokonta. There are thousands of different fungi species sharing our environment, exposing human to them in many ways including the air we breathe, the food we consume and even the water we drink. There is a significant number of fungi that could cause infections in immunodeficient individuals and in individuals with genetic variants that promote susceptibility to specific fungal organisms. Fortunately, methods for isolation of specific groups of fungi are well defined and developed allowing researchers in generating knowledge, conceptual understanding and insight regarding the kingdom fungi itself. The first group of fungi that will be focused on is the Endophytic fungi, also called plant hidden fungi, defined as fungi which part or all of their life cycle living naturally in the plant tissues without causing disease symptoms. Endophytic fungi can be isolated from the leaves, stems, flowers, fruits, and seeds. *Serendipita indica*, is one example of root endophytic fungi that has numerous applications in agriculture. Another group of fungi that will also be introduced in this practical is aquatic fungi. Aquatic fungi are microscopic organisms with mostly mycelial growth and hyphae developing on or within their typically submerged organic substrates of plant or animal origin. They are usually found in stagnant ponds, ditches, or slow flowing water and are capable of vegetative growth on submerged leaves and woody substrates under semi-anaerobic conditions. *Paraphysoderrma sedebokerense* is one of the fungi species from phylum Blastocladiomycota that lives in freshwater habitat. It is a facultative pathogenic chytrid that causes irreversible damage to some green microalgae.

Objectives:

1. To isolate unknown fungi species from different sources (example: leaves, stems, fruits, ditches, slow flowing water, etc.)
2. To grow unknown fungi species on suitable growth medium.
3. To identify fungi species by using Olympus CX12 light microscope.

Bioprocess Technology Lab Manual

Michael C. Flickinger

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analyze samples in forensic clinical quality control environmental and other testing laboratories Lab Manual for Herren's Introduction to Biotechnology, 2nd Ray V. Herren,2011-12-30 The Laboratory Manual is a valuable tool designed to enhance your lab experience and give you an opportunity to experience hands on the materials covered in the core text Lab activities objectives materials lists step by step procedures illustrations and review questions are found in the Lab Manual Plant Biotechnology: Laboratory Manual For Plant Biotechnology Chawla,2004 This practical laboratory manual has been designed to familiarise students with protocols on plant tissue culture and recombinant DNA technology It deals with the basic aspects on introduction laboratory organization sterilization techniques nutrition medium and the choice of explant It also has exercises on plant tissue culture seed culture embryo culture meristem culture node culture axillary bud proliferation etc A part of the manual also deals with recombinant DNA technology **Biochemistry and Biotechnology** ,2014 **Lab**

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A Laboratory Manual Of Plant Biotechnology (2Nd Ed.) S. S. Purohit, 2004

Molecular Biology Techniques Susan Carson, Heather B. Miller, Sue Carson, D. Scott Witherow, 2011-11-07 This manual is an indispensable tool for introducing advanced undergraduates and beginning graduate students to the techniques of recombinant DNA technology or gene cloning and expression. The techniques used in basic research and biotechnology laboratories are covered in detail. Students gain hands on experience from start to finish in subcloning a gene into an expression vector through purification of the recombinant protein. The third edition has been completely re-written with new laboratory exercises and all new illustrations and text designed for a typical 15 week semester rather than a 4 week intensive course. The project approach to experiments was maintained; students still follow a cloning project through to completion, culminating in the purification of recombinant protein. It takes advantage of the enhanced green fluorescent protein; students can actually visualize positive clones following IPTG induction. Cover basic concepts and techniques used in molecular biology research labs. Student tested labs proven successful in a real classroom laboratories. Exercises simulate a cloning project that would be performed in a real research lab. Project approach to experiments gives students an overview of the entire process. Prep list appendix contains necessary recipes and catalog numbers providing staff with detailed instructions.

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